

Electronic Journal of Biotechnology

journal homepage: www.elsevier.com/locate/ejbt

Short Communication

Draft genome of Pseudomonas sp. RGM 2987 isolated from Stevia philippiana roots reveals its potential as a plant biostimulant and potentially constitutes a novel species

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GRAPHICAL ABSTRACT

ARTICLE INFO

Article history: Received 27 April 2022 Revised 28 September 2022 Accepted 4 October 2022 Available online 13 October 2022

Keywords: Biostimulant Chilean Altiplano Draft genome Plant growth-promoting bacteria Pseudomonas Pseudomonas novel species Rhizobacteria Rhizosphere Roots Stevia philippiana

Background: Reports on Pseudomonas species associated with Chilean native plants and their diversity are scarce. The draft genome of Pseudomonas sp. RGM 2987 isolated from the rhizosphere of Stevia philippiana, a native Chilean plant from the Altiplano, is presented.

Results: The assembled genome features 6,161,133 bp, a G+C content of 61.3%, and 5,350 predicted open reading frames. dDDH, and ANIb differences between RGM 2987 and the closest relatives support its classification as a new species within the Pseudomonas genus. Genome mining and functional classification revealed the presence of genes involved in cell function and metabolism as well as plant-growth promotion, including those for indole acetic acid production, phosphate solubilization, and ethylene concentration.

Conclusions: The draft genome of Pseudomonas sp. RGM 2987 provides insights on its phylogeny and classification as a new species and shed light on its potential as a plant-biostimulant, expanding our knowledge on Pseudomonas biodiversity worldwide and, specifically, for those strains associated with native Chilean plants.

How to cite: Guerra M, Carrasco-Fernández J, Valdés JH, et al. Draft genome of Pseudomonas sp. RGM 2987 isolated from Stevia philippiana roots reveals its potential as a plant biostimulant and potentially constitutes a novel species. Electron J Biotechnol 2022;61. https://doi.org/10.1016/j.ejbt.2022.10. 001.

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Peer review under responsibility of Pontificia Universidad Católica de Valparaíso ⇑ Corresponding author.

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1. Introduction

Pseudomonas is a diverse genus of aerobic, gram-negative bacteria, present in diverse habitats, including plant rhizosphere and

<https://doi.org/10.1016/j.ejbt.2022.10.001>

0717-3458/ 2022 Pontificia Universidad Católica de Valparaíso. Production and hosting by Elsevier B.V. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>). endosphere, and contains more than 200 validly published species [\[1\]](#page-4-0). Species of the *Pseudomonas* genus are known for exhibiting plant-growth promotion traits, such as indole acetic acid (IAA) biosynthesis, stress alleviation enzyme production, and phosphate solubilization, which are desirable properties for the development of effective plant biostimulants [\[2\].](#page-4-0)

Chile is one of the top five plant biodiversity hotspots on the planet [\[3\]](#page-4-0); however, little information regarding Pseudomonas species associated with native Chilean plants can be found in the literature. Pseudomonas atacamensis is one of the formally described species isolated from the rhizosphere of desert bloom plants in the Atacama region, and its draft genome revealed important plant growth-promoting (PGP) traits including iron acquisition and alleviation of abiotic stress $[4,5]$. Here, we present the draft genome of Pseudomonas sp. RGM 2987, isolated from the rhizosphere of Stevia philippiana, a native Chilean plant, distributed from Arica-Parinacota to Atacama regions in northern Chile and is a relative of the worldwide known Stevia rebaudiana, source of the natural sweetener "stevia" $[6]$. We explored its taxonomic affiliation as a novel species and its biotechnological potential as a plant biostimulant.

2. Experimental

2.1. Bacteria isolation

A plant sample of S. philippiana was collected in 2017 from the Chilean Altiplano at Pachama, Putre, Arica-Parinacota region, Chile (-18.454733, -69.510400; altitude: 3,400 m.a.s.l.), placed inside a sterile plastic bag, and transported in a dark container to the Microbial Genetic Resources Bank at INIA. Rhizosphere soil was removed from the roots by soaking them in an epiphyte removal buffer $[7]$. Next, 100 µL of serial dilutions of the soil rhizosphere of S. philippiana were inoculated on King's B (KB) agar medium supplemented with 25 μ g/mL nystatin and 50 μ g/mL cycloheximide. Plates were incubated at 25° C for 48 h. A UV-fluorescent colony was streaked out on KB agar and incubated at 25° C for 48 h. This step was repeated twice to obtain an axenic culture. The isolate was deposited in the Chilean Collection of Microbial Genetic Resources (CChRGM) under the code RGM 2987.

2.2. In vitro assessment of plant growth promotion traits

RGM 2987 was grown in Luria Bertani broth to prepare a bacterial suspension of 10^8 CFU/mL in sterile distilled water. IAA production was evaluated by inoculating 20 mL YMD broth, supplemented with 1% L-tryptophan, with 10 μ L of the cell suspension. The culture was incubated for 72 h at 25° C and 120 rpm. IAA was quantified according to Carrasco-Fernández et al. [\[8\].](#page-4-0) Phosphate solubilization was evaluated by inoculating the center of a Petri dish containing NBRIP agar with 10 µL of the cell suspension. Plates were incubated at 28° C for 7 d. A clear halo around the colony was considered as positive solubilization of phosphate, and the phosphate solubilization index was calculated according to Demissie et al. [\[9\].](#page-4-0)

2.3. Isolation of genomic DNA, 16S rRNA gene amplification, and whole-genome sequencing

Genomic DNA was isolated using the Wizard[®] Genomic DNA Purification Kit (Promega, USA), following the Gram-negative strain protocol. The 16S rRNA gene sequencing was conducted according to Carrasco-Fernández et al. [\[8\]](#page-4-0). DNA extraction and

library construction was performed at MicrobesNG (UK), according to the provider's methodology [\[10\]](#page-4-0). Libraries were sequenced using Illumina sequencers (HiSeq/NovaSeq) using a 250-bp paired-end protocol.

2.4. Bioinformatic analyses

Nucleotide sequences from Sanger sequencing were processed using Sequencher 5.4.6 (Gene Codes Corporation, USA). EzBioCloud was used for 16S rRNA gene identification [\[11\].](#page-4-0) Whole-genome sequencing reads were adapter trimmed with Trimmomatic 0.30 using a sliding window quality cutoff of Q15 [\[12\]](#page-4-0). De novo assem-bly was performed using SPAdes 3.7 [\[13\]](#page-4-0), and contigs were annotated with the Prokaryotic Genome Annotation Pipeline (PGAP) 6.0 [\[14\]](#page-4-0). The Conserved Domain Database was used for protein domain search [\[15\].](#page-4-0) Digital DNA-DNA hybridization (dDDH) and 16S rRNA gene phylogeny (maximun parsimony and maximun likelihood trees were evaluated with 1000 bootstrap iterations) were performed in the Genome-to-Genome Distance Calculator [\[16\]](#page-4-0) webserver. The Average Nucleotide Identity using BLAST (ANIb) calculation was performed with JSpeciesWS [\[17\]](#page-4-0). Reference genomes were retrieved from GenBank [\[18\]](#page-4-0). ANIb values were used for Pearson correlation matrix construction, which were subsequently ordered by hierarchical clustering with the R package cor-rplot in R [\[19\]](#page-4-0). Comparison of the RNA polymerase sigma factor (rpoD) nucleotide sequence of RGM 2987 was performed against a dataset of rpoD sequences from reference Pseudomonas strains [\[1\]](#page-4-0). Search for particular genes within the genome annotation was performed using local BLAST analysis on Sequenceserver [\[20\]](#page-4-0) and functional Clusters of Orthologous Groups (COGs) of proteins were obtained from Global Catalogue of Type Strain (gcType) [\[21\]](#page-4-0).

3. Results and discussion

3.1. Genome sequence features and taxonomic analysis of Pseudomonas sp. RGM 2987

The RGM 2987 genome was 6,161,133 bp in length (223 contigs), with 61.3% G+C content, mean coverage of 31.79x, and N_{50} of 59,539; 5,350 open reading frames and 60 tRNAs were predicted. According to the similarities of the 16S rRNA gene, RGM 2987 belonged to the genus Pseudomonas, sharing the highest similarity with Pseudomonas mucoides $P154a^T$ (98.86%). The maximum likelihood phylogenetic tree separates the strain RGM 2987 as a new branch in the Pseudomonas phylogeny and diverges from its closest relatives with 100% branch support [\(Fig 1A](#page-2-0)). dDDH values support 16S rRNA gene phylogeny, indicating that RGM 2987 corresponds to a new species (cutoff $\leq 70\%$ [\[22\]](#page-4-0)) as well as the ANIb values (cutoff \leq 95% [\[23\]](#page-4-0)) and their clustering patterns within the Pseudomonas representatives [\(Fig 1B\)](#page-2-0). According to the rpoD gene similarities, the strain RGM 2987 was grouped within members of the Pseudomonas corrugata subgroup, being Pseudomonas zanjanensis SWRI12^T, a strain isolated from wheat rhizosphere, its closest relative [\[1\]](#page-4-0).

3.2. In vitro assessment of plant growth promotion features

The strain RGM 2987 produced 41.05 ± 7.68 µg/mL IAA and displayed a phosphate solubilization index of 1.55 \pm 0.09 [\(Fig 1C\)](#page-2-0), which suggests its potential use as a plant-growth promoting rhizobacteria [\[8,24\]](#page-4-0).

Fig. 1. Taxonomic, genomic, and phenotypic features of Pseudomonas sp. RGM 2987. (a) Maximum likelihood (ML) tree of 16S rRNA gene sequences inferred under the GTR +GAMMA model and rooted by midpoint-rooting, scale bar indicates substitutions per site. The numbers above the branches are support values when larger than 60% from ML (left) and maximum parsimony (right) bootstrapping; (b) correlation plot based on ANIb values between each species (species name is indicated with the respective key in the y-axis). ANIb and dDDH values from pairwise whole-genome comparisons between Pseudomonas sp. RGM 2987 and the closest type strains are indicated at the bottom; (c) clear zone (halo) produced by RGM 2987; (d) COG features retrieved from the RGM 2987 genome sequence.

Table 1

Plant-growth promotion traits found in Pseudomonas sp. RGM 2987.

-, no significant hit was found.

3.3. Biotechnological potential of Pseudomonas sp. RGM 2987 as a plant biostimulant

The genome annotation of strain RGM 2987 revealed the presence of multiple open reading frames for multiple cellular function, secondary metabolism, and other of unknown function ([Fig 1D\)](#page-2-0); PGP traits found in the genome are summarized in Table 1. Phosphate-solubilizing bacteria are capable of solubilizing phosphate present in soil by releasing organic acids, of which gluconic acid is mostly produced by them [\[25\]](#page-4-0). Bacteria produce gluconic acid by the direct oxidation of glucose through the quinoprotein glucose dehydrogenase enzyme, encoded by the gcd gene [\[26\].](#page-4-0) Gcd uses pyrroloquinoline quinone as a cofactor, which is pro-duced by the activities encoded in the pqqA-F operon [\[27\]](#page-4-0). The gcd gene and the six genes comprising the pqq operon were identified in RGM 2987, implying the presence of a well-conserved mechanism for phosphate solubilization in soil through gluconic acid synthesis.

IAA is an auxin hormone involved in plant growth and development. It can be synthesized by the plant itself and by soil bacteria, and its production levels may depend upon tryptophan availability and abiotic factors [\[28,29\].](#page-4-0) Soil bacteria can synthesize IAA from Ltryptophan by multiple interconnected pathways, such as: (i) indole-3-pyruvate (IPyA), (ii) indole-3-acetamide (IAM), and (iii) indole-3-acetonitrile (IAN) [\[29,30\]](#page-4-0). A key enzyme for IAA biosynthesis in the IPyA pathway is the indole-3-pyruvate decarboxylase enzyme, encoded by the ipdC gene. It belongs to the indol_phenyl_DC superfamily (cl37262) and its disruption in Azospirillum brasilense reduced \sim 50% IAA production [\[31\].](#page-4-0) RGM2987_05960 was the closest ortholog to IpdC in strain RGM 2987; however, it lacks the domain cl37262. Two enzymes participate in the IAM pathway, a tryptophan 2-monooxygenase (IaaM) and an indole acetamide hydrolase (IaaH). Pseudomonas sp. RGM 2987 contains two ortholog proteins of IaaM, of which only RGM2987_16825 belongs to the same superfamily as IaaM (cl38049), of the plant pathogenic bacteria Pseudomonas savastanoi [\[32\]](#page-4-0). No homologues for the iaaH gene were detected in the genome of RGM 2987 suggesting that the IAM route may be truncated in this strain. Two sub-routes to synthesize IAA in the IAN pathway have been described: one generates IAA from the IAN substrate through a reaction catalyzed by a nitrilase enzyme encoded by the nit gene, and a second route transforms IAN into IAM intermediate by a nitrile hydratase, encoded by nthAB genes [\[33\]](#page-4-0). A nitrilase encoding gene was found in strain RGM 2987, which belonged to the nitrilases_CHs superfamily (cd07564) as well as the Nit protein of Pseu-domonas sp. UW4 [\[33\].](#page-4-0) No nhtAB genes were found in RGM 2987

suggesting that IAA production in RGM 2987 may occur through a nitrilase enzyme in the IAN pathway.

The molecule 1-aminocyclopropane-1-carboxylate (ACC), the precursor of ethylene in higher plants, is a gaseous phytohormone that governs plant development and, depending on its concentration, it may inhibit or induce plant senescence under a wide range of stress conditions $[34,35]$. The gene acdS encodes for an ACC deaminase that participates in the conversion of ACC into ammonia and α -ketobutyrate [\[36\].](#page-4-0) The presence of *acdS* and its regulatory gene, acdR, in the genome of RGM 2987 suggest a potential modulatory mechanism for ethylene concentration.

4. Conclusions

The genome of Pseudomonas sp. RGM 2987 revealed important plant-growth promoting traits such as a route for IAA biosynthesis, phosphate solubilization, and ethylene concentration regulation, which are desirable features for developing plant-biostimulants. Phylogenetic and phylogenomic analysis provide strong support to claim that the strain RGM 2987 represents a novel species of the Pseudomonas genus, thus expanding the knowledge regarding microbial biodiversity associated with native plants in Chile.

Author contribution

Study conception and design: JF Castro; M Guerra

Data collection: J Carrasco-Fernández

Genome sequencing and data analysis: M Guerra; JH Valdés

Analysis and interpretation of results: JF Castro, JH Valdés, M Panichini, M Guerra

Draft manuscript preparation: JF Castro; M Guerra; J Carrasco-Fernández; JH Valdés

Revision of the results and approved the final version of the manuscript: JF Castro; JH Valdés

Financial support

The authors acknowledge funding from Núcleos de Investigación (Grant: 502953-70) of the Chilean National Agency for Research and Development (INIA), Chile and to the FONDEQUIP Program from the ANID (Grant: EQM200205) for funding a platform of equipment for the preservation of microbial genetic resources.

Conflict of interest

None.

Data availability

This Whole Genome Shotgun project has been deposited at DDBJ/ENA/GenBank under the accession [JALJDX000000000.](https://www.ncbi.nlm.nih.gov/nuccore/JALJDX000000000) The version presented here is [JALJDX000000000.1.](https://www.ncbi.nlm.nih.gov/nuccore/JALJDX000000000) Raw reads have been deposited in the Sequence Read Archive (SRA) under the accession numbers SRR18649284 and SRR18649285. All project data are available under BioProject accession number [PRJNA820732.](https://www.ncbi.nlm.nih.gov/bioproject/?term=PRJNA820732) 16S rRNA and rpoD gene sequences were submitted to the NCBI GenBank under the accession numbers [OP364341](https://www.ncbi.nlm.nih.gov/nuccore/OP364341) and [OP380681,](https://www.ncbi.nlm.nih.gov/nuccore/OP380681) respectively.

References

- [1] Girard L, Lood C, Höfte M, et al. The ever-expanding Pseudomonas genus: description of 43 new species and partition of the Pseudomonas putida group.
Microorganisms 2021;9(8):1766. https://doi.org/10.3390/ [https://doi.org/10.3390/](https://doi.org/10.3390/microorganisms9081766) [microorganisms9081766.](https://doi.org/10.3390/microorganisms9081766) PMid: 34442845.
- [2] Compant S, Cambon M, Vacher C, et al. The plant endosphere world bacterial life within plants. Environ Microbiol 2020;23(4):1812-29. [https://doi.org/](https://doi.org/10.1111/1462-2920.15240) [10.1111/1462-2920.15240](https://doi.org/10.1111/1462-2920.15240). PMid: 32955144.
- [3] Vargas R, Reif A, Faúndez MJ. The forests of Robinson Crusoe Island, Chile: an endemism hotspot in danger. Bosque (Valdivia) 2011;32(2):155–64. [https://](https://doi.org/10.4067/S0717-92002011000200006) [doi.org/10.4067/S0717-92002011000200006.](https://doi.org/10.4067/S0717-92002011000200006)
- [4] Poblete-Morales M, Carvajal D, Almasia R, et al. Pseudomonas atacamensis sp. nov., isolated from the rhizosphere of desert bloom plant in the region of Atacama, Chile. Antonie van Leeuwenhoek 2020;113(8):1201–11. [https://doi.](https://doi.org/10.1007/s10482-020-01427-0) [org/10.1007/s10482-020-01427-0](https://doi.org/10.1007/s10482-020-01427-0). PMid: 32436126.
- [5] Poblete-Morales M, Plaza N, Almasia R, et al. Draft genome sequence of Pseudomonas sp. strain M7D1, isolated from the rhizosphere of desert bloom plants. Microbiol Resour Announc 2019;8(22):e00441–e519. [https://doi.org/](https://doi.org/10.1128/MRA.00441-19) [10.1128/MRA.00441-19.](https://doi.org/10.1128/MRA.00441-19) PMid: 31147434.
- [6] Gutiérrez DG, Muñoz-Schick M, Grossi MA, et al. The genus Stevia (Eupatorieae, Asteraceae) in Chile: a taxonomical and morphological analysis. Phytotaxa 2016;282(1). [https://doi.org/10.11646/phytotaxa.282.1.1.](https://doi.org/10.11646/phytotaxa.282.1.1)
- [7] Simmons T, Caddell DF, Deng S, et al. Exploring the root microbiome: extracting bacterial community data from the soil, rhizosphere, and root endosphere. JoVE 2018;135:e57561. <https://doi.org/10.3791/57561>. PMid: 29782021.
- [8] Carrasco-Fernández J, Guerra M, Castro JF, et al. Plant growth promoting rhizobacteria from Juan Fernández archipelago improve germination rate of endangered plant Solanum fernandezianum Phil. Chilean J Agric Res 2020;80 (1):41–9. [https://doi.org/10.4067/S0718-58392020000100041.](https://doi.org/10.4067/S0718-58392020000100041)
- [9] Demissie S, Muleta D, Berecha G. Effect of phosphate solublizing bacteria on seed germination and seedling growth of faba bean (Vicia faba L.). Int. J Agric Res 2013;8(3):123–36. <https://doi.org/10.3923/ijar.2013.123.136>.
- [10] MicrobesNG. MicrobesNG sequencing service methods v20210419, 2021 [cited 08 March 2022]. Available from Internet: [microbesng.com/documents/24/MicrobesNG_Sequencing_Service_Methods_](https://microbesng.com/documents/24/MicrobesNG_Sequencing_Service_Methods_v20210419.pdf) [v20210419.pdf.](https://microbesng.com/documents/24/MicrobesNG_Sequencing_Service_Methods_v20210419.pdf)
- [11] Yoon S-H, Ha S-M, Kwon S, et al. Introducing EzBioCloud: a taxonomically united database of 16S rRNA gene sequences and whole-genome assemblies. Int J Syst Evol Microbiol 2017;67(5):1613–7. [https://doi.org/10.1099/](https://doi.org/10.1099/ijsem.0.001755) isem.0.001755. PMid: 28005526.
- [12] Bolger AM, Lohse M, Usadel B. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 2014;30(15):2114–20. [https://doi.org/10.1093/](https://doi.org/10.1093/bioinformatics/btu170) [bioinformatics/btu170](https://doi.org/10.1093/bioinformatics/btu170). PMid: 24695404.
- [13] Bankevich A, Nurk S, Antipov D, et al. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 2012;19 (5):455–77. [https://doi.org/10.1089/cmb.2012.0021.](https://doi.org/10.1089/cmb.2012.0021) PMid: 22506599.
- [14] Li W, O'Neill KR, Haft DH, et al. RefSeq: expanding the Prokaryotic Genome Annotation Pipeline reach with protein family model curation. Nucleic Acids Res 2021;49(D1):D1020–8. [https://doi.org/10.1093/nar/gkaa1105.](https://doi.org/10.1093/nar/gkaa1105) PMid: 33270901.
- [15] Marchler-Bauer A, Derbyshire MK, Gonzales NR, et al. CDD: NCBI's conserved domain database. Nucleic Acids Res 2015;43(D1):D222-6. [https://doi.org/](https://doi.org/10.1093/nar/gku1221) [10.1093/nar/gku1221.](https://doi.org/10.1093/nar/gku1221) PMid: 25414356.
- [16] Meier-Kolthoff JP, Carbasse JS, Peinado-Olarte RL, et al. TYGS and LPSN: a database tandem for fast and reliable genome-based classification and nomenclature of prokaryotes. Nucleic Acids Res 2022;50(D1):D801–7. [https://doi.org/10.1093/nar/gkab902.](https://doi.org/10.1093/nar/gkab902) PMid: 34634793.
- [17] Richter M, Rosselló-Móra R, Oliver Glöckner F, et al. JSpeciesWS: a web server for prokaryotic species circumscription based on pairwise genome comparison. Bioinformatics 2016;32(6):929–31. [https://doi.org/10.1093/](https://doi.org/10.1093/bioinformatics/btv681) [bioinformatics/btv681.](https://doi.org/10.1093/bioinformatics/btv681) PMid: 26576653.
- [18] Sayers EW, Cavanaugh M, Clark K, et al. GenBank. Nucleic Acids Res 2021;49 (D1):D92–6. [https://doi.org/10.1093/nar/gkaa1023.](https://doi.org/10.1093/nar/gkaa1023) PMid: 33196830.
- [19] R Core Team. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. 2018 [cited 08 March 2022]. Available from Internet: [https://www.R-project.org/.](https://www.R-project.org/)
- [20] Priyam A, Woodcroft BJ, Rai V, et al. Sequenceserver: a modern graphical user interface for custom BLAST databases. Mol Biol Evol 2019;36(12):2922–4. <https://doi.org/10.1093/molbev/msz185>. PMid: 31411700.
- [21] Shi W, Sun Q, Fan G, et al. gcType: a high-quality type strain genome database for microbial phylogenetic and functional research. Nucleic Acids Res 2020;49 (D1):D694–705. <https://doi.org/10.1093/nar/gkaa957>. PMid: 33119759.
- [22] Meier-Kolthoff JP, Göker M, Spröer C, et al. When should a DDH experiment be mandatory in microbial taxonomy? Arch Microbiol 2013;195(6):413–8. org/10.1007/s00203-013-0888-4. PMid: 23591456
- [23] Richter M, Rosselló-Móra R. Shifting the genomic gold standard for the prokaryotic species definition. Proc Natl Acad Sci USA 2009;106 (45):19126–31. [https://doi.org/10.1073/pnas.0906412106.](https://doi.org/10.1073/pnas.0906412106) PMid: 19855009.
- [24] Kalayu G. Phosphate solubilizing microorganisms: promising approach as biofertilizers. Int J Agron 2019;2019:4917256. [https://doi.org/10.1155/2019/](https://doi.org/10.1155/2019/4917256) [4917256.](https://doi.org/10.1155/2019/4917256)
- [25] Sharma SB, Sayyed RZ, Trivedi MH, et al. Phosphate solubilizing microbes: sustainable approach for managing phosphorus deficiency in agricultural soils. SpringerPlus 2013;2(1):587. <https://doi.org/10.1186/2193-1801-2-587>. PMid: 25674415.
- [26] Suleman M, Yasmin S, Rasul M, et al. Phosphate solubilizing bacteria with glucose dehydrogenase gene for phosphorus uptake and beneficial effects on wheat. PLoS ONE 2018;13(9):e0204408. [https://doi.org/10.1371/journal.](https://doi.org/10.1371/journal.pone.0204408) [pone.0204408](https://doi.org/10.1371/journal.pone.0204408). PMid: 30240432.
- [27] Rasul M, Yasmin S, Suleman M, et al. Glucose dehydrogenase gene containing phosphobacteria for biofortification of Phosphorus with growth promotion of rice. Microbiol Res 2019;223–225:1–12. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.micres.2019.03.004) [micres.2019.03.004](https://doi.org/10.1016/j.micres.2019.03.004). PMid: 31178042.
- [28] Lambers H, Chapin III FS, Pons TL. Growth and Allocation. In: Plant Physiological Ecology. New York, NY: Springer; 2008. p. 321-74. [https://doi.](https://doi.org/10.1007/978-0-387-78341-3_10) [org/10.1007/978-0-387-78341-3_10.](https://doi.org/10.1007/978-0-387-78341-3_10)
- [29] Duca DR, Rose DR, Glick BR. Indole acetic acid overproduction transformants of the rhizobacterium Pseudomonas sp. UW4. Antonie van Leeuwenhoek 2018;111(9):1645–60. [https://doi.org/10.1007/s10482-018-1051-7.](https://doi.org/10.1007/s10482-018-1051-7) PMid: 29492769.
- [30] Spaepen S, Vanderleyden J, Remans R. Indole-3-acetic acid in microbial and microorganism-plant signaling. FEMS Microbiol Rev 2007;31(4):425–48. [https://doi.org/10.1111/j.1574-6976.2007.00072.x.](https://doi.org/10.1111/j.1574-6976.2007.00072.x) PMid: 17509086.
- [31] Malhotra M, Srivastava S. An ipdC gene knock-out of Azospirillum brasilense strain SM and its implications on indole-3-acetic acid biosynthesis and plant growth promotion. Antonie van Leeuwenhoek 2008;93(4):425-33. [https://doi.](https://doi.org/10.1007/s10482-007-9207-x) [org/10.1007/s10482-007-9207-x](https://doi.org/10.1007/s10482-007-9207-x). PMid: 17952626.
- [32] Yamada T, Palm CJ, Brooks B, et al. Nucleotide sequences of the Pseudomonas savastanoi indoleacetic acid genes show homology with Agrobacterium tumefaciens T-DNA. Proc Natl Acad Sci USA 1985;82(19):6522–6. [https://doi.](https://doi.org/10.1073/pnas.82.19.6522) [org/10.1073/pnas.82.19.6522](https://doi.org/10.1073/pnas.82.19.6522). PMid: 16593610.
- [33] Duca D, Rose DR, Glick BR, et al. Characterization of a nitrilase and a nitrile hydratase from Pseudomonas sp. strain UW4 that converts indole-3- acetonitrile to indole-3-acetic acid. Appl Environ Microbiol 2014;80 (15):4640–9. <https://doi.org/10.1128/AEM.00649-14>. PMid: 24837382.
- [34] Iqbal N, Khan NA, Ferrante A, et al. Ethylene role in plant growth, development and senescence: interaction with other phytohormones. Front Plant Sci 2017;8:475. [https://doi.org/10.3389/fpls.2017.00475.](https://doi.org/10.3389/fpls.2017.00475)
- [35] Orozco-Mosqueda MC, Duan J, DiBernardo M, et al. The production of ACC deaminase and trehalose by the plant growth promoting bacterium Pseudomonas sp. UW4 synergistically protect tomato plants against salt stress. Front Microbiol 2019;10:1392. [https://doi.org/10.3389/](https://doi.org/10.3389/fmicb.2019.01392) [fmicb.2019.01392.](https://doi.org/10.3389/fmicb.2019.01392) PMid: 31275294.
- [36] Glick BR, Nascimento FX. Pseudomonas 1-aminocyclopropane-1 carboxylate (ACC) deaminase and its role in beneficial plant-microbe interactions. Microorganisms 2021;9(12):2467. [https://doi.org/10.3390/](https://doi.org/10.3390/microorganisms9122467) [microorganisms9122467.](https://doi.org/10.3390/microorganisms9122467) PMid: 34946069.